VIOMYCIN I. THE AMINO ACID SEQUENCE OF VIOMYCIN Tsunehiro Kitagawa, Yosuke Sawada, Takako Miura, Teruaki Ozasa and Hyozo Taniyama

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Many attempts have been made to elucidate the structure of viomycin, a strongly basic tuberculostatic streptomyces antibiotic (1-7). These studies led Bowie, Cox, Johnson and Thomas (6) to present I and more recently Dyer, Kellogg, Nassar and Streetman (7) to propose II as the structure of viomycin. This report describes our determination of the amino acid sequence of viomy-



Complete acid hydrolysis of viomycin yielded the amino acids L-serine, L-Q, β -diaminopropionic acid (dapa), L- β -lysine and viomycidine^{*1}(ratio: 2:1:1:1) and urea, carbon dioxide and ammonia, as reported by others(5,6).

Partial hydrolysis of viomycin was achieved with 1 N hydrochloric acid at 100⁰ for six hours. The hydrolysates were separated by column chromatography on active charcoal, eluting with water, 5% acetic acid and 5% acetic acid-20% phenol. From the water eluate L-serine and urea were obtained.

The 5% acetic acid eluate was rechromatographed on an Amberlite CG-120 column and β -lysine and peptide III isolated, [d]_n²⁷⁰-2° (0.1% in H₂°);

^{*1} Amino acid analyse were carried out, using an Hitachi Amino acid analyser KLA-3, with 50 cm. Column.

Rf*2: 0.6; R*3:0.8; no ultraviolet absorption; positive Sakaguchi reaction.

The 5% acetic acid-20% phenol eluate was chromatographed on a cellulose powder column and 3 peptides isolated: Peptide Ia, $[\mathbf{C}]_{D}^{27^{\circ}}$ -43° (1% in H₂O); Rf: 0.25; R: 0.98; λ max 269.5 m μ (log ε 3.21); positive Sakaguchi reaction. Peptide Ib, $[\mathbf{C}]_{D}^{12^{\circ}}$ -49.5°(1% in H₂O); Rf: 0.11; R: 0.88; λ max 285 m μ (log ε 3.93); positive Sakaguchi reaction. Peptide II, $[\mathbf{C}]_{D}^{17^{\circ}}$ -38°(1% in H₂O); Rf: 0.5; R: 0.95; λ max 268 m μ (log ε 4.12); positive Sakaguchi reaction.

Dyer <u>et</u>. <u>al</u>. (7) reported the hydrolysis of viomycin with 0.1 N hydrochloric acid at 90° for 8 hours to give urea and desureaviomycin. Following the same hydrolysis and chromatographing the hydrolysates on an active charcoal column we obtained urea ^{*4} and peptide Ic^{*5}, [**d**]_D^{16°}-37° (1% in H₂O); Rf: 0; R: 0.9; λ max 272 m µ (log £ 3.95); positive Sakaguchi reaction.

Peptide III gave L-serine, L-dapa and viomycidine on complete hydrolysis. Hydrazinolysis of peptide III gave viomycidine as a free amino acid. Hydrolysis of the <u>bis</u>-2,4-dinitrophenyl peptide III (λ max 365 m μ , log ϵ 4.44) yielded viomycidine, 2,4-dinitrophenylserine and <u>mono</u>-2,4-dinitrophenyldapa.

For comparison, the two possible <u>mono-</u>2,4-dinitrophenyldapa acids were synthesized by the modified copper complex method (8) from L-dapa, which was made from L-aspartic acid by a Schmidt reaction (9). Thus, we obtained \mathbf{q} -amino- β -2,4-dinitrophenylamino propionic acid, m.p.205°; anal. calcd. for $C_9H_{10}N_4O_6$: C, 40.00; H, 3.55; N, 20.74; found: C, 39.54; H, 3.55; N, 20.08; $[\mathbf{q}]_2^{25^\circ}$ + 77.11° (0.82% in 0.2 N HCI); and the hygroscopic β -amino- \mathbf{q} -2,4-dinitrophenylaminopropionic acid from acid hydrolysis of β -benzoylamino- \mathbf{q} -2,4-dinitrophenylaminopropionic acid m.p. 236-238°; anal. calcd. for $C_{16}H_{14}N_4O_7$: c, 51.34; H, 3.77; N, 14.65; found: C, 51.61; H, 3.77; N, 14.65.

^{*2} Rf values were obtained on PPC with Toyoroshi No.51 paper and a solvent of (t-BuOH:HOAc: H₂O, 2:1:1) for development.

^{*3} Electrophoresis was carried out with a Toyo-C type (500V, 3-5 mA) instrument for 3-5 hrs. with Toyoroshi No.51 paper and pyridine: HOAc:H₂0,(5:0.2:95) (pH 6.5) buffer solution. R values were obtained with reference to viomycin defining the electophoresis distance of viomycin as 1, under the above conditions.

^{*4} Urea was detected by the urease method (10).

^{*5} The amino acid components, N-terminal amino acid and C-terminal amino acid of peptide Ic are identical to those of desureaviomycin which was obtained by Dyer <u>et. al.(7)</u>. They report that desureaviomycin has no ultraviolet absorption, but the peptide Ic has ultraviolet absorption. c.f. Ultraviolet absorvance of viomycin was followed in O.1 N HCI at 100°. The ultraviolet absorvance decreased gradually and after 6 hrs. it had disapeared. But a lyophilized product of this solution showed ultraviolet absorption.

The <u>mono-2</u>,4-dinitrophenyl dapa obtained from peptide III could be assigned the q -amino- β -2, 4-dinitrophenylaminopropionic acid structure by comparison of its Rf and R values with those of the corresponding synthetic compound: Rf₁ : 0.76 (n-butanol: acetic acid: water, 2:1:1) and Rf₂: 0.53 (n-butanol : t-butanol : pyridine: acetic acid : water, 15:4.5:10:3:12) on PPC and R₁: 0.47 (ethanol 20 ml. containing 20 drops of acetic acid) and R₂ : 0.53 (Phenol: ethanol: water, 15:4:1) on TLC using Kieselgel G. In contrast the isomeric β -amino - q - 2,4-dinitrophenylaminopropionic acid showed Rf₁: 0.82; Rf₂: 0.74; R₁: 0.20; R₂: 0.14.

Thus, the amino acid sequence of peptide III is determined as seryl-dapayl-viomycidine, where the β -amino group of dapa is free.

On complete acid hydrolysis peptide II and each member of the peptide I group (Ia, Ib, Ic) gave L-serine, L-dapa, L- β -lysine and viomycidine (ratios 2:1:1:1) respectively.^{*6} The same amino acids were obtained from the acid hydrolysis of viomycin.

Hydrazinolysis of viomycin did not give a free amino acid, *7 but hydrazinolysis of peptide II and of the peptide I group (Ia, b, c) gave L-serine as a free amino acid.

Acid hydrolysis of the <u>bis</u>-2,4-dinitrophenyl derivatives of viomycin and of the peptide I group gave all of the amino acids described above except β -lysine. <u>Bis</u>-2,4-dinitrophenyl- β -lysine was obtained in 80% yield from the viomycin derivative. Similarly it was obtained from each of the peptide I group derivatives and identified by PPC and TLC.

Acid hydrolysis of the <u>tri</u>-2,4-dinitrophenylpeptide II ($\lambda \max 370 \text{ m} \mu$, logg 4.31) gave free serine and vicmycidine and the derivatives <u>bis</u>-2,4-dinitrophenyl- β -lysine and **q** -amino- β -2,4dinitrophenylaminopropionic acid. Therefore, the amino acid sequence of peptide II is:

 β -lysyl-seryl-diamino(- β -NH₂)propionyl-viomycidyl-serine where the β amino group of dapa is free and the amino acid sequence of the peptide I group is:

 β -lysyl-seryl-dapayl(β -NH-)-viomycidyl-serine

where the β -amino group of dapa is tied up. From these results one concludes that the amino acid sequence of viomycin is :

 β -lysyl-seryl-dapayl(- β -NH-)-viomycidyl-seryl-urea III

where the β -amino group of dapa is not free. This sequence is not consistent with structure I, proposed by Bowie <u>et</u>. <u>al</u>.(5,6) or with structure II, proposed by Dyer <u>et</u>. <u>al</u>. (7) for viomycin.

The first group obtained on partial hydrolysis of viomycin with 6 N hydrochloric acid a pep-

^{*6} The amino acid sequence of the peptide I group are identical but each peptide shows different ultraviolet absorption and different Rf values on PPC and R values on electrophoresis. Each peptide shows a single spot on PPC in different solvent systems and on electrophoresis.

^{*7} The same result was obtained by Dyer et. al. (7).

tes of a 2,4-dinitrophenyl derivative of viomycin yielded <u>mono-</u> q-2,4-dinitrophenyldapa and <u>mono-</u> q-2,4-dinitrophenyl- β -lysine. From these results they deduce the amino acid sequence and present structure I for viomycin.

Dyer <u>et</u>. <u>al</u>. (7) pointed out that hydrolysis of <u>bis</u>-2,4-dinitrophenylviomycin gave only <u>bis</u>-2,4-dinitrophenyl- β -lysine, consistent with our results. In addition they reported that they obtained serine and viomycinic acid from the action of carboxypeptidase on desureaviomycin (c.f. *5) and postulate a serine-serine-urea unit in viomycin. On treatment of viomycin with water they obtained a compound (which gave serine and lysine on hydrolysis) which they presumed to correspond to peptide B of Bowie <u>et</u>. <u>al</u>. (6) but for which they proposed a seryl ester of β -lysine.

From these results and the structure of peptide A obtained by Bowie <u>et</u>. <u>al</u>. the amino acid sequence was deduced and structure II proposed for viomycin.

Examining the action of carboxypeptidase A ^{*8} on peptide Ic, which we assume to be identical with the above desureaviomycin, we could not obtain serine nor viomycinic acid. Since we could demonstrate the seryl-dapaylviomycidine group in viomycin and since viomycin contains two seryl units, one can deduce the presence of a viomycidyl-seryl-urea unit and exclude a viomycidyl-seryl-seryl-urea unit. Thus the amino acid sequence in viomycin must be that shown in III.

For the structure of viomycidine Dyer <u>et</u>. <u>al</u>. (4) presented IV which was supported by Bowie <u>et</u>. <u>al</u>. (5) but subsequently changed to V (7).



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Bowie <u>et</u>. <u>al</u>. (6) reported that their peptide A shows ultraviolet absorption at 275 m μ (£ 5100) and assigned structure VI to it whereas Dyer <u>et</u>. <u>al</u>. (7) presented VII for peptide A. These

*8 Bought from Sigma chemical company.

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structures were incorporated into the proposals I and II for viomycin.



Both structures VI and VII are not expected to show the positive Sakaguchi reaction found with viomycin and peptide A. Structures I and II also fail to explain the instability of the chromophore of viomycin (c.f.*5). The structure of the chromophore of viomycin is currently under investigation. An explanation for the release of the urea unit from viomycin under very mild conditions will be presented later.

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